

REMARKS

Reconsideration is respectfully requested.

Claims 1-50, 66, 70, 73, and 89 have been cancelled. Claims 51-65, 67-69, 71, 72, 74-88, and 90-93 are pending. Applicants also have filed a request for continued examination and supplemental information disclosure statement with this response.

With respect to all amendments and cancelled claims, Applicant has not dedicated or abandoned any unclaimed subject matter and moreover have not acquiesced to any rejections and/or objections made by the Patent Office. Applicant reserves the right to pursue prosecution of any presently excluded claim embodiments in future continuation and/or divisional applications.

Withdrawn Rejections and Objections

Applicants respectfully thank the Examiner for withdrawing the rejections and objections in the office action dated December 12, 2005.

Declaration Under 37 C.F.R. § 1.131

The Examiner objected to the declaration under 37 C.F.R. § 1.131 previously submitted on July 14, 2004 because the declaration was unsigned. Applicants herewith resubmitted a signed declaration in accordance with 37 C.F.R. § 1.131, as discussed below.

Information Disclosure Statement.

The Examiner states that reference C10 was not submitted to the Office.

Applicants herewith include a Supplemental Information Disclosure Statement including reference C10. Applicants respectfully request its consideration.

Claim Interpretation

The Examiner states that “the term ‘electrode’ will be considered as meaning ‘a solid support comprising a metallic surface.’”

The Examiner also states that “the term ‘shielding’ is used here in its everyday meaning, i.e. ‘preventing physical contact.’ Therefore, ‘blocking moieties shielding nucleic acids from the electrode’ means any structural elements which prevent contact target and/or probe nucleic acids with the electrode.”

Applicants respectfully traverse these interpretations. Applicants do not take a position with respect to the interpretation, as no such position is necessary given the accompanying declaration.

35 USC § 102(e)

Claims 51-58, 60-62, 64-73, 79, 80, 82, 83, 85-89 and 93 stand rejected under 35 USC § 102(e) as anticipated by Wohlstadter et al., US Patent No. 6,066,448 ("Wohlstadter"). Wohlstadter is a continuation-in-part of USSN 08/402,076, filed March 10, 1995, which is a continuation-in-part of USSN 08/402,277, also filed March 10, 1995. Accordingly, the earliest possible priority date available for the disclosure in Wohlstadter is March 10, 1995.

The Applicants herewith submit a Declaration under 37 C.F.R. §1.131 by inventors Thomas J. Meade and Jon F. Kayyem. The Declaration demonstrates that the claimed invention was made prior to the earliest possible March 10, 1995 priority date of Wohlstadter.

To antedate a 35 USC 102(e) reference, the inventors must show possession of a species within a claimed genus. As stated in the MPEP:

[t]he 37 CFR 1.131 affidavit or declaration must establish possession of either the whole invention claimed or something falling within the claim (such as a species of a claimed genus), in the sense that the claim as a whole reads on it. *In re Tanczyn*, 347 F.2d 830, 146 USPQ 298 (CCPA 1965). MPEP 715.02.

The inventors need not show possession subject matter identical to that of the references. As further stated in the MPEP:

a 37 CFR 1.131 affidavit is not insufficient merely because it does not show the identical disclosure of the reference(s) or the identical subject matter involved in the activity relied upon. If the affidavit contains facts showing a completion of the invention commensurate with the extent of the invention as claimed is shown in the reference or activity, the affidavit or declaration is sufficient, whether or not it is a

showing of the identical disclosure of the reference or the identical subject matter involved in the activity.
See *In re Wakefield*, 422 F.2d 897, 164 USPQ 636 (CCPA 1970). MPEP 715.02.

The Declaration and associated Exhibits demonstrate that the claimed invention was made prior to the earliest Wohlstadter priority date. Paragraph 5 of the Declaration summarizes an embodiment of the invention within the scope of claim 51.

The Declaration discloses the production of an array of claim 51 depicted in Exhibit A. Different regions on the array are defined by 8x8 micron squares on the photolithographic mask. The gold surface is the electrode of claim 51. The thiol-(CH₂)₁₆-OH is the blocking and linking moiety. When the thiol-(CH₂)₁₆-OH is covalently attached to the nucleic acid and the gold surface, it is the linking entity. When the thiol-(CH₂)₁₆-OH is attached to the gold surface, and not attached to the nucleic acid, it is the blocking moiety.

The fluorescent complement is an agent that distinguishes between single stranded and double stranded nucleic acids. Dark squares indicate locations where single stranded nucleic acids were ablated off, and light squares indicate where nucleic acid hybrids were present. A montage of images is depicted in Exhibit A.

The declaration outlines that the invention was completed in this country prior to March 10, 1995. Because the claimed invention was made prior to the earliest Wohlstadter priority date, the Wohlstadter reference is not prior art. Applicants respectfully request withdrawal of this ground for rejection.

35 USC § 103(a)

Claims 59, 63, 81, 84 and 85 stand rejected under 35 USC § 103(a) as being unpatentable over Wohlstadter in view of Kayyem et al., U.S. Patent No. 6,096,273 (Kayyem).

In a separate rejection, claims 75-78 and 90-92 also stand rejected 35 USC § 103(a) as being unpatentable over Wohlstadter in view of Kayyem.

As demonstrated above in the response to the rejection under 35 USC § 102(e), Wohlstadter is not a prior art reference. Therefore, Wohlstadter cannot be combined with Kayyem as described by the Examiner in satisfaction of the requirements of 35 USC § 103(a). Accordingly, Applicants respectfully request withdrawal of this ground for rejection.

CONCLUSION

On the basis of the amendments and remarks presented herein, Applicants believe that this application is in a condition of allowance. Applicants respectfully request that the Examiner pass this application to issue, and early notification of such is requested.

If the Examiner has any questions, she is invited to call the undersigned at (415) 781-1989.

Respectfully submitted,

DORSEY & WHITNEY LLP

Dated: May 20, 2005 By: [Signature]

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Filed Under 37 C.F.R. § 1.34



PATENT

Attorney Docket No.: A-64411-2

Attorney File No.: 468267-00067

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of

MEADE *et al.*

Serial No.: 09/921,645

Filed: August 3, 2001

For: *Metallic Solid Supports Modified
with Nucleic Acids*

Examiner: STRZELECKA, Teresa, E.

Group No. 1637

"EXPRESS MAIL" LABEL NO.:
EV 554099109 US

DECLARATION PURSUANT TO 37 C.F.R. § 1.131

Mail Stop RCE
Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313

Sir:

We, Thomas J. Meade and Jon F. Kayyem hereby declare as follows:

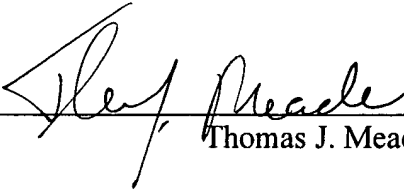
1. We are the inventors on the above-identified patent application and are familiar with its contents. We have also reviewed the pending claims in this application.
2. We are familiar with the Office Action mailed on November 17, 2003 wherein claims 51-69, 71, 72, and 74-93 were rejected over Wohlstadter et al. (6,066,448) which has an earliest possible priority date of March 10, 1995.
3. All of the ideas detailed in the above-identified application were contemplated in this country prior to March 10, 1995. This is evidenced by the appended documents.
4. One of the goals of the project that led to the filing of the parent application was to create a surface comprising a self-assembled monolayer with single stranded nucleic acids attached (referred herein as probes), and then to answer three questions: first, whether a solution-based complementary strand would bind to the probe; second, would a complementary strand attached to an atomic force

microscopy (AFM) tip bind to the probe, and if so; third, whether or not we could determine the force necessary to “tear apart” the duplex.

5. The experiments started out with the synthesis of the monolayer portion using an HO-(CH₂)₁₆-OH to form a molecule with a protected sulfur group (for attachment to a gold surface) on one end, to which a phosphoramidated nucleic acid was attached. The experiments proceeded with the coating of a gold surface with this monolayer-forming material. A photolithographic mask, with 8 x 8 micron squares on it, was then used to cover the gold surface. The surface was then exposed to a photoactivated agent and a mercury arc lamp which resulted in the ablation nucleic acids from the squares not covered by the mask. We then added a fluorescent complement to the surface, and viewed it under a confocal microscope. This resulted in a pattern of “light”, e.g. fluorescent, background, where the fluorescent solution based probes were found, and “dark” squares, where the surface-bound single stranded nucleic acid had been ablated off, and therefore no fluorescent probe was detected. A montage of several of these images, made over the course of the experiments, is attached as Exhibit A.
6. With regard to timing of these experiments, the documents attached as Exhibit B are pages from the notebook detailing the synthesis of some of the compounds used in these experiments. (Please note that all experiments not relevant to the present discussion have been redacted, as have all dates.) For example, page 136 documents the conversion of the HO-(CH₂)₁₆-OH molecule to the asymmetrical HO-(CH₂)₁₆-OAc needed for further reactions. The bottom of page 139 and the top of page 140 show the synthesis of the protected thiol-(CH₂)₁₆-OH molecule. the top of page 141 shows the reaction of the protected thiol-(CH₂)₁₆-OH molecule added to a phosphoramidite moiety. In conclusion, the invention was completed in this country prior to March 10, 1995.
7. We declare further that all statements made herein of our own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that the making of willful false statements and the like are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such

willful statements may jeopardize the validity of the application or any patent
issuing thereon.

Date: 5-4-05


Thomas J. Meade

Date: _____

Jon F. Kayyem

willful statements may jeopardize the validity of the application or any patent
issuing thereon.

Date: _____

Thomas J. Meade

Date: May 18, 2005


Jon F. Kayyem

EXHIBIT A

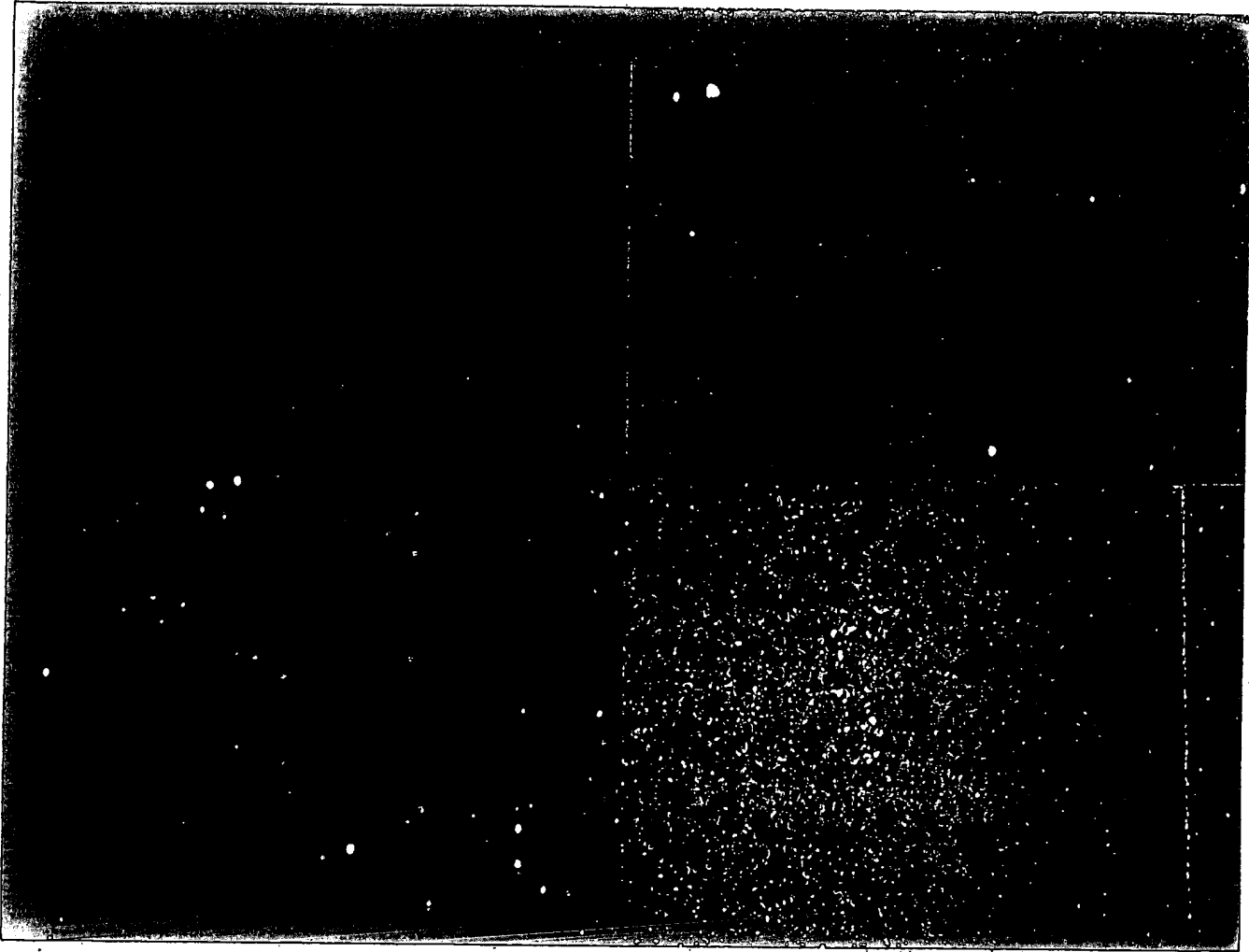
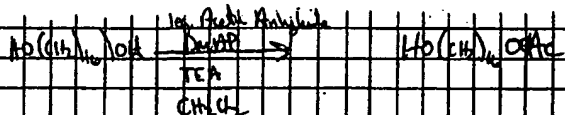
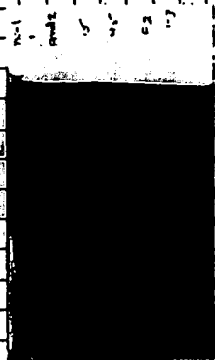


EXHIBIT B



0.5 gr of $\text{HO(CH}_2\text{)}_n\text{OH}$ (m.w. 258.45; 1.94×10^{-3} moles) was placed in a small round bottom and 20 ml of CH_2Cl_2 added along with 0.05 equiv (9750⁻⁵ moles 12.7 or 11.8 mg) and 1.4 equiv of TEA (4.7 ml) and 1 equiv of Arctic Anhydride (m.w. 102.9; d = 1.02) (10.5 ml or 1.73 ml).



25.50 gms/lb

→ continue to next

12.7 mg
 11.8 mg

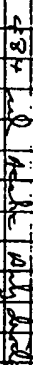
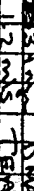
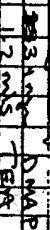
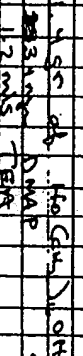
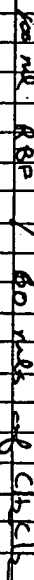
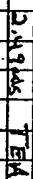
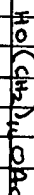
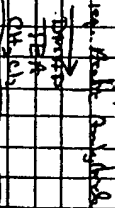
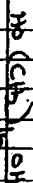
10/20
 10/20

Rec. 2

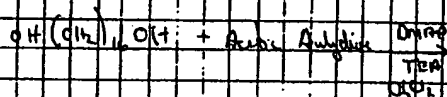
2.05 gr (7.93×10^{-3} moles) was placed in a 100 ml RBF and 60 ml of CH_2Cl_2 added along with 0.05 equiv DMAP (4.8 mg) and 1.4 equiv of TEA (1.7 ml) and 6.95 ml of Arctic Anhydride.

2

150

[illegible]

Rec B



1.57 g of Diol were dissolved in 25 ml of dry CH_2Cl_2 . 0.5 g of DMAP (32.33 mg) was added and 1.4 eq of TEA (1.4 ml) and 4% of Acetic Anhydride

Tube 72

by mass double

20 ml solvent from 7 ml diol
12 mg new sample = 2.5
35 mg new sample = 1.4 eq

The flash column was

20 250 ml of silica

1.2 liters of 180:20 hex/ether

was passed through and the
gradient pushed up to 80

55:45. After tube 72

it broke and 200 ml of 55:45

to remove all the product

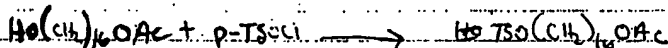
→ 72

→ new sample

→ Diol 150:50 Ether/hexane

 $\text{C}_{18}\text{H}_{34}\text{O}_2$

360.53



Excerpt from page 1180

500mg (1.7 x 10⁻³ moles) of $\text{HO}(\text{CH}_2)_{16}\text{OCC(=O)CH}_3$ in in dry pyridine and in 50 ml ethylmethyl ether and cooled to 0°C. p-TsCl (10.65) in 1 ml ether excess (or 634 mg) with a string bar and allowed to proceed for 90 hrs. The solution was poured into a beaker with 200 ml of ice water and Section filtered. The solid slurred in hexane (20) and returned to

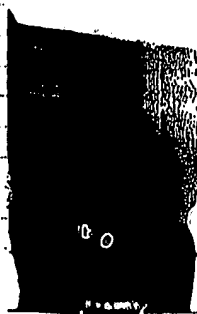
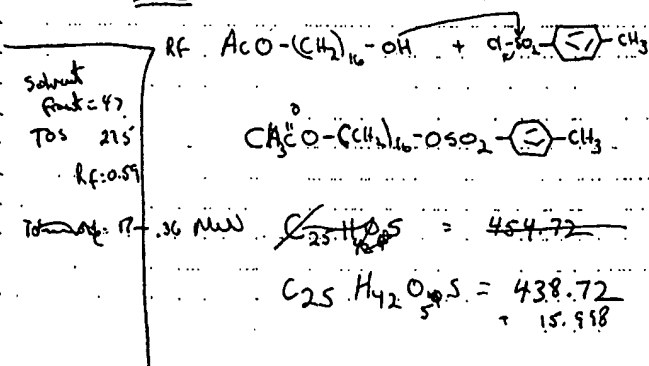
| Spot | Rf |
|-------|------|
| Start | 0 |
| Prod | 0.25 |
| Tos | 0.58 |

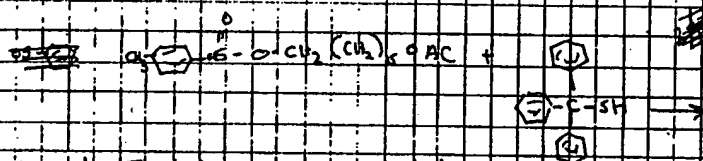


p-TsCl
The product spot is UV active and chem active. It is probably the p-TsO in the C18M32. The p-TsO is also UV active (+) active.

degrees. H¹NMR is consistent with the proposed structure

Repeat





1370 mg of TSO₄ was dissolved in 10 ml of DNF and thoroughly digested on the vac. line ($\eta_{sp}/c = 454.72$ or 8.4×10^{-4} ml/dm³)

WJF-40

- Triphenyl methyl reagent ($MW = 270.30$, $0.95 \text{ equiv} = 7.7 \times 10^{-2}$ moles)
[$= 213 \text{ mg}$]
- 31.8 mg of NaOH ($.78 \text{ equiv}$) in more of H_2O

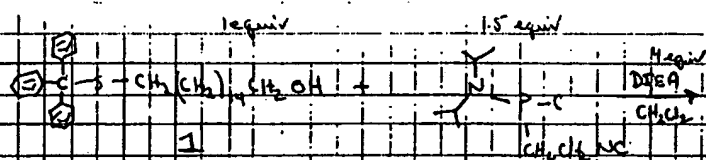
5 ml of ethanol was degassed on the vac line and 213 mg of triethylamine added. 10.180 ml of degassed NaOEt in Et_2O was added via syringe under Ar . To this solution 370 mg of $\text{TSD}(\text{CH}_3)_6\text{PF}_6$ in DMF/EtOH was added and the solution degassed.

After treatment with NaOH in micell \rightarrow

solvent = 49mm
 int. lig. = 0.4
 = 19mm
 or 86:14

.84,-

The material was taken up in MeOH/MeOH sol


 $\text{C}_{24}\text{H}_{46}\text{O}_5$ - MW 504.87

• 220 mg of 1 = 4.4×10^{-4} moles

• 6.5×10^{-4} moles of phosphoramidite 236.68 $d = 1.061$ $M = 4.48$ ml

5.74 ml • 2.0×10^{-4} moles DIEA MW 125.25 $\frac{0.792}{125.25} = 5.74 \times 10^{-4}$ moles

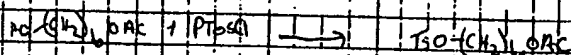
220 mg of 1 was stirred in 15 ml of dry CH_2Cl_2 and immediately 320 μl of DIEA was added. 145 μl of cyanateyl phosphoramidite was added dropwise. After some and additional 50 μl of cyanateyl reagent was added.

Note:

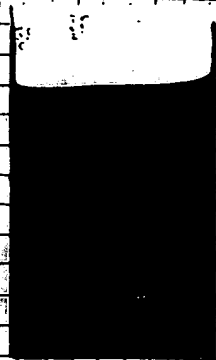
TEA MUST be present during flash!

• At the addition of 1% TEA to the mobile phase, swap the eluent

Tris-04



57 gr of monac product was stirred in 350 ml of dry pyridine and cooled to 0°C. 650 mg of TosCl was added and the reaction mixture allowed to proceed @ 40 hrs.

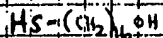
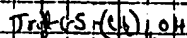


Note:

Must use FRESH TBocI

Total yield 150 mg

50/50 EtAc/Hexane



Starting 11mg of Tris-S-(CH₃)₆OH (2.13 moles)

276.40

275.39 g

516.89

3 mmol

100 mM TEAE

100 mM AgNO₃

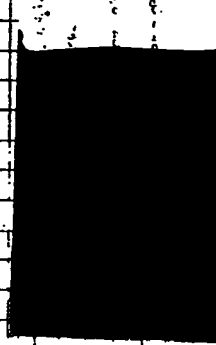
140 mM DTT

1 mg dissolved in MeOH (5 ml) and adding TEAE buffer.

1 ml of AgNO₃ solution - 30 min

1 ml of DTT + 30 min (154.2 mg) 152.5 mg/ml = 1 M

1 ml of 100 mM DTT or 1 ml of 1 M DTT
1 ml of 100 mM AgNO₃ or 1 ml of 1 M AgNO₃



NMR says NR. The majority of isolated material is strongly reduced.

$$\text{TSO}-(\text{CH}_2)_n-\text{OAc} + \text{NaSH} \xrightarrow{\text{DMF}}$$

| | |
|----------|--|
| 16 equiv | |
| 2 equiv | |

5

10

$$m_w = 454.72$$

от 3.3 до 4
от 18.4 до 19

| | |
|----|--------|
| or | 18.4 m |
|----|--------|

NaI + Anhydrous
Acetone

Entire page

NMR reveals that the spot identified ① is from the reaction of triethyl-OH with AgNO_3 did not occur

NMR = triple OH NH

50:50

Doc
After: 1/1/19

$$\approx 75 \text{ mgs starting material} = 1.7 \times 10^{-4} \text{ moles} \times 1.2 \text{ equiv of DMSH}$$

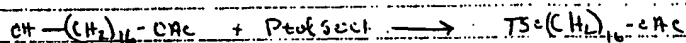
2×10^{-4} moles NaOH or 8 mg dissolved in MeOH

901 Mono AC

5/20/92

50:60
dist / time

$x = y \cos \theta$



2 x 250mg or 8.5×10^{-4} moles in 15 ml of dry pyridine is

cooled to 0°C in an ice bath. 320 mgs of TSOCl is added

and the reaction allowed to proceed @ 4°C . The product (brownish tint) is poured into a beaker with amounts of

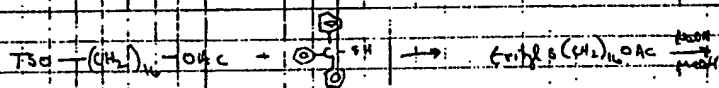
ice water, stirred for 10 min and filtered. The solid is

washed with water and dissolved in pet ether and

charcoal, added with stirring. The mixture was filtered

and dried (vacuum totd)

24 hours is not long enough, some oil remains



325 mgs (mw = 454.72) or 7.15×10^{-4} moles was dissolved in 10 ml of

dry DMF and degassed. Triphenylmethylisocyanate (mw 276.4) with

1.1 equiv (7.86×10^{-4}) or 217 mgs and 1.05 equiv of NaOH or 30 mgs

was dissolved in 150 ml of H_2O and degassed.

The triphyl SH was dissolved in 5 ml of EtOH and degassed. The NaOH

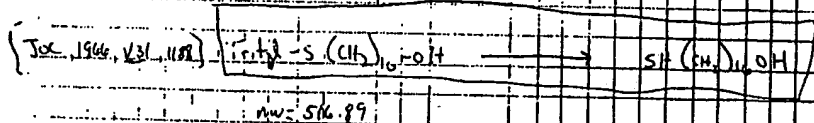
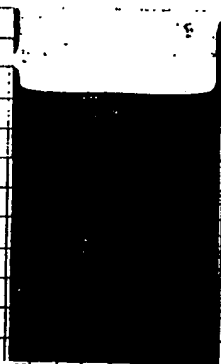
was added via syringe and then the TSO-OAc added. The reaction

was repeatedly degassed and allowed to proceed for 12 hrs.

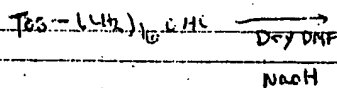
The reaction was TLC, and NaOH/MeOH added to check for

⇒ Flash → phosphoric acid

EtOAc



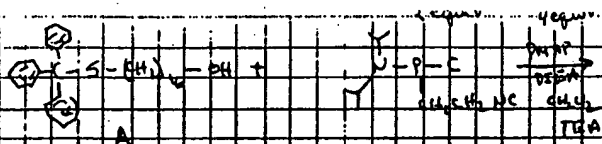
25 mg (4.8×10^{-2} mols) were dissolved in ^{small} acetic acid (0.15 ml) and
 39 mls of 1N HCl added. The reaction was allowed to stir for 15
 hrs at 75°C. Upon addition of 100 ml, the mixture formed a white
 ppt immediately. After warming for ~ 5 min the ppt redissolved
 with the resulting solution slightly cloudy.



MW = 96.72

0.45 g of the Tos derivative was dissolved in 5 ml of dry DMF.
 1.4×10^{-4} moles of 2 equiv of NaOH or 2.9×10^{-4} moles of 17
 equiv of NaOH is dissolved in 1 ml of dry MeOH. The NaOH
 (in 2 ml of dry MeOH) was degassed and the MeOH added.
 This solution is added (via syringe) to the dry DMF Tos
 product.

148.



- A plate using the Ammonium Cerium Nitrate revealed on the
presence of DTT

After cooling a white ppt. formed and the solution was
dissolved in 100 ml of CH_2Cl_2 . $pH \approx 10$ (pH 9.5)
was used to wash the CH_2Cl_2 . The reaction was
continued until the water layer was pH 8. The solution
was then washed with pH 7 buffer, returned to
original

5

11

→ Before base wash

↳ After washing (neutralized) with pH 9 Na_2CO_3

poly-d-lysine (mw 25,000) + DTPA Anhydride →

Polylysine was dissolved in D_2O and (250 mg in 2 ml) and applied to a PD-10. An additional 1/2 ml was added and 3 ml total collected from each of 3 PD-10 columns. The recovered dried material (≈ 65 mg) was divided into 3 reaction vessels of 22 mg each.

The following ratios will be prepared:

50x
100x
200x

(See page 100 for details)

0.1 mm poly-d-lysine:

22 mg (8.6×10^{-6} moles) in $\boxed{8.6 \text{ ml}}$ = 0.1 mM

50x DTPA Anhydride

or $(4.3 \times 10^{-5}) \times 357.3 = 15.4 \text{ mg}$

100x

or $(8.6 \times 10^{-5}) \times 357.3 = 30.7 \text{ mg}$

200x

or $(1.72 \times 10^{-4}) \times 357.3 = 61.5 \text{ mg}$

0.5 M HCO_3^- buffer pH 9.5

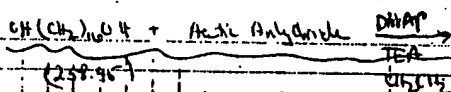
Only the 200x reaction showed any change in pH (e.g. $\rightarrow 9.4$)

The reaction was ~~extremely~~ rapid, as usual to lysine

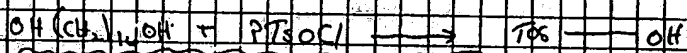
400x

35.2 mg or 1.4×10^{-6} moles

200 mg of DTPA Anhydride in 14 ml pH 9.5 ~~area~~
Buffer

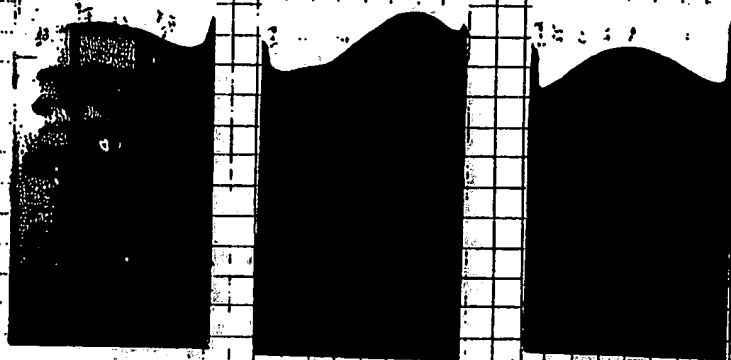


3 gr of diol was stirred in 50 ml of CH_2Cl_2 and
~~2.5~~ 7 mm of DMAP added along with 2.5 ml of TEA
 and 1.04 ml of Acetic Anhydride



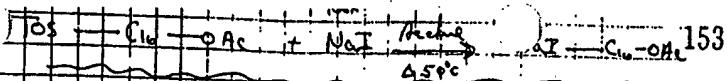
2 gr of $\text{CH(CH}_3\text{)}_2\text{OH}$ (7.74×10^{-3} moles) ^{sample} was dissolved in 30 ml
 of dry pyridine and cooled to 0°C . TsCl (190.65) required
 = 1.49 grams was added and the reaction allowed to proceed
 for

TOS — C16 — OAc Purification



The water/py solution is expected to degenerate and should be
heptane/dichloromethane and evaporated 2 hrs. The solid is
dissolved in 50% 2-propanol and filtered. The solution is
applied to a column 45:5 2-propanol:EtOAc. Peak 2 is visible
immediately. Future: Run 90:10 to start and run up to 80:20

See page 135 for prep.



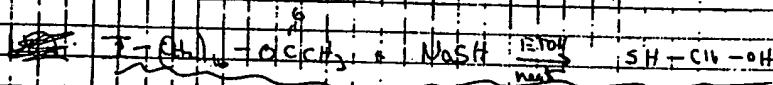
180 mg

150

120

50 mg of $\text{Ios} - \text{OAc}$ ($\text{MW} = 454.72$) or 5.1×10^{-4} moles is dissolved in
 100 ml Acetone and a 100% excess of 6.2×10^{-4} moles of NaI (MW 149.89)

of NaI added. The reaction is followed by TLC. After 15 min
 a crystalline material begins to put out



50 mg of $\text{I} - (\text{CH}_2)_4 - \text{OAc}$ was
 dissolved in 1.5 ml of EtOH.

75 mg of NaSH (10x) was

shaken in 10 ml of EtOH

and the iodide added slowly

and kept at -10°C for 2 hrs.

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